

- 1 R.A. Robinson, in: *Plant Pathosystems*, p.184. Springer-Verlag, Berlin 1976.
- 2 J.H. Andrews, *Biol. Rev.* 51, 211 (1976).
- 3 J.H. Andrews, *Can. J. Bot.* 55, 1019 (1977).
- 4 Y. Nozawa and K. Nozawa, *Bull. Jap. Soc. scient. Fish.* 23, 427 (1957).
- 5 G.N. Agrios, in: *Plant Pathology*, 2nd edn, p.703. Academic Press, New York 1978.
- 6 J.M. Sieburth, *Adv. Microbiol. Sea I*, 63 (1968).
- 7 K.M. Khailov and Z.P. Burlakova, *Limnol. Oceanogr.* 14, 521 (1969).
- 8 T.R. Parsons, in: *The Ecology of the Seas*, p.81. Ed. D.H. Cushing and J.J. Walsh. W.B. Saunders Co., Philadelphia 1976.
- 9 J. Kohlmeyer, *Mar. Biol.* 8, 344 (1971).
- 10 J.E. Van Der Plank, in: *Plant Diseases: Epidemics and Control*, p.349. Academic Press, New York 1963.
- 11 J.H. Ryther, in: *Preliminary Results with a Pilot Plant Waste Recycling Marine-Aquaculture System*, p.50. WHOI-75-41, Woods Hole, Massachusetts.
- 12 J.S. Prince, *Aquaculture* 4, 69 (1974).

SPECIALIA

The editors do not hold themselves responsible for the opinions expressed in the authors' brief reports. – Les auteurs sont seuls responsables des opinions exprimées dans ces brèves communications. – Für die Kurzmitteilungen ist ausschliesslich der Autor verantwortlich. – Per le brevi comunicazioni è responsabile solo l'autore. – Ответственность за короткие сообщения несёт исключительно автор. – Solo los autores son responsables de las opiniones expresadas en estas comunicaciones breves.

Sesquiterpenes based on the cadalane skeleton from the brown alga *Dilophus fasciola*¹

V. Amico, G. Oriente, M. Piattelli, C. Tringali, E. Fattorusso, S. Magno and L. Mayol

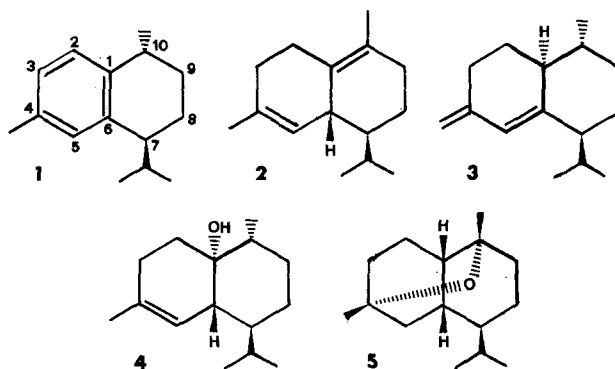
Istituto di Chimica Organica dell'Università, viale Doria 8, I-95125 Catania (Italy), and Istituto di Chimica Organica dell'Università, via Mezzocannone 16, I-80134 Napoli (Italy), 17 July 1978

Summary. From the brown alga *Dilophus fasciola* (Dictyotaceae) the new sesquiterpene ether 4,10-epoxymurolane has been isolated, along with known sesquiterpenes based on the cadalane skeleton, and its structure determined by spectroscopic methods.

The algal family Dictyotaceae is a rich source of terpenoids and other metabolites². In a recent paper³ we reported the isolation of a compound of mixed biogenesis, namely geranylgeranyl glycerol, from the more polar fractions of the chloroform extract of *Dilophus fasciola* (Roth) Howe, a brown alga belonging to this family. An investigation of the less polar fractions of the same extract has now led to the isolation of 5 sesquiterpenoids based on the cadalane skeleton, the hydrocarbons 1–3, the alcohol 4 and the novel cyclic ether 5.

Material and methods. *D. fasciola* (2.5 kg fresh weight) was collected near Catania, Sicily, during the spring of 1977. The chloroform extract of the freeze-dried alga was subjected to a gross fractionation on silica gel using increasing concentrations of ether in hexane as the eluent. From the early fractions of the hexane eluent, a hydrocarbon mixture was obtained, wherefrom isolation of individual components in pure form required extensive use of chromatography over AgNO₃ impregnated silica gel. Later eluates, which emerged from the column with 10% ether, on further chromatography over silica gel using benzene as eluent, gave the alcohol 4 and the ether 5, and in addition 2 further oxygenated sesquiterpenoids currently under investigation. Aromatization was performed by heating at 250 °C for 3 h the pertinent compound (100 mg) with 10% Pd/C (100 mg). The crude product was chromatographed on silica gel (hexane as eluent) to give cadalene, identified by comparison of physical properties (UV, IR and NMR) with those of a reference sample. Partial aromatization of 1-epibicyclosesquiphellandrene was carried out with trifluoroacetic acid in the condition described by Andersen et al.⁴.

Results and discussion. For all the isolated compounds the cadalane skeleton was established by dehydrogenation to cadalene. Compound 1 (0.1% dry weight of the alga) C₁₅H₂₂, and compound 2 (0.02%), C₁₅H₂₄, were identified, respectively as (1S-trans)-(-)-calamenene and δ -cadinene by comparison of their physical data ([α]_D, MS, UV, IR and NMR) with those reported in the literature^{4,5}. The 3rd hydrocarbon (0.25%), C₁₅H₂₄, [α]_D = +4.1° (c 1 in EtOH), was a conjugated diene that had spectral properties which matched those reported for 1-epibicyclosesquiphellandrene⁶. However, since the OR of this compound, recently isolated from *Ocimum basilicum*⁶, has not been recorded, the possibility that the algal metabolite could be its enantiomer was taken into consideration but definitely ruled out in view of the formation of (1S-trans)-(-)-calamenene by partial aromatization of 3.



The alcohol **4**, $C_{15}H_{26}O$ (0.03%), was identified as cubenol by comparison of $[\alpha]_D$ and spectral data with those reported in the literature^{5,7}. Compound **5** was isolated as an oil (0.08%), $[\alpha]_D = +15.4^\circ$ (c 1 in EtOH). High resolution mass spectrometry established the elemental composition as $C_{15}H_{26}O$. The IR- and UV-spectra indicated the absence of hydroxyl or carbonyl group in the molecule. The single oxygen atom must be part of an ether bridge connecting 2 fully substituted carbon atoms; the IR-spectrum exhibited strong absorption at 1090 cm^{-1} , while the ^{13}C -NMR showed 2 singlets at 72.45 and 69.65 ppm. This spectrum also comprised 4 methines (47.27, 32.36, 30.89, 28.04), 5 methylenes (39.33, 34.80, 33.96, 27.31, 18.86) and 4 methyls (25.91, 22.29, 22.29, 21.34). The ^1H -NMR spectrum (270 MHz, CDCl_3) displayed singlets at δ 1.12 and 1.14 assignable to tertiary methyls attached to oxygen-bearing carbons. Other diagnostically valuable signals appeared at δ 0.69 (3H, d, $J=6.7\text{ Hz}$) and 0.93 (3H, d, $J=6.7\text{ Hz}$) and were assigned to methyls of an isopropyl group, since they collapsed to singlets by irradiation at frequency of an 1H multiplet at δ 1.73. From the above data it was established that **5** possessed the cadalane skeleton bearing an ether bridge connecting position 4 and 10. Closure of the oxane ring requires a cis relationship between H-1, H-6, Me-4 and Me-10. The relative stereochemistry of the remaining chiral centre at C-7 was established by the following criteria. a) Irradiation at δ 0.91 caused the isopropyl methine multiplet to collapse into a slightly broadened singlet; this revealed that the 1H signal masked by methyl groups, but evidenced by integration, was due to H-7. b) The signal of this proton and those of the isopropyl group remained almost unaffected by the addition of $\text{Eu}(\text{fod})_3$ and thus the isopropyl-bearing ring must have, as expected, a chair conformation. c) In the ^1H -NMR spectrum a signal (dddd, $J=13.5, 13.5, 4.5, 4.5$) is seen at δ 1.98, partially obscured by other protons, but well separated in C_6D_6 ; this signal, which suffers a remarkable europium shift, is simplified to

a double-double doublet ($J=13.5, 13.5, 4.5$) by irradiation at the frequency of H-7 (0.91); this result can only be explained assuming that the signal at δ 1.98 is due to the axial proton attached to C-8 and that H-7 is equatorial. Therefore, the new ether is 4,10-epoxymurolane possessing the relative stereochemistry depicted in **5**. It is worth noting that *D. fasciola* accumulates sesquiterpenoids based on the cadalane skeleton, whereas the congener species *D. ligulatus* synthesizes perhydroazulene diterpenes⁸. Cadalane sesquiterpenes have been previously isolated from algae of the related genus *Dictyopteris* (cadalene, $(-)$ - γ_1 -cadinene and $(-)$ - δ -cadinol from *D. divaricata*⁹ and zonarene from *D. zonaroides*)¹⁰.

- 1 This work was carried out in the frame of the 'Progetto finalizzato per l'Oceanografia e i Fondi marini', C.N.R., Rome, Italy.
- 2 J.T. Baker and V. Murphy, Compound from marine organisms, vol. I. CRC Press, Cleveland 1976; P.J. Scheuer, Chemistry of marine natural products. Academic Press, New York 1973.
- 3 V. Amico, G. Oriente, M. Piattelli, C. Tringali, E. Fattorusso, S. Magno and L. Mayol, *Experientia* 33, 989 (1977).
- 4 N.H. Andersen, D.D. Syzdal and C. Graham, *Tetrahedron Lett.* 1972, 905.
- 5 B.A. Nagasampagi, L. Yankov and Sukh Dev, *Tetrahedron Lett.* 1968, 1913.
- 6 S.J. Terhune, J.W. Hogg and B.M. Lawrence, *Phytochemistry* 13, 1183 (1974).
- 7 Y. Ohta and Y. Hirose, *Tetrahedron Lett.* 1967, 2073.
- 8 V. Amico, G. Oriente, M. Piattelli, C. Tringali, E. Fattorusso, S. Magno and L. Mayol, *J. chem. Soc. chem. Commun.* 1976, 1069; B. Danise, L. Minale, R. Riccio, V. Amico, G. Oriente, M. Piattelli, C. Tringali, E. Fattorusso, S. Magno and L. Mayol, *Experientia* 33, 413 (1977).
- 9 T. Irie, K. Yamamoto and T. Masamune, *Bull. chem. Soc. Japan* 37, 1053 (1964).
- 10 W. Fenical, J.J. Sims, R.M. Wing and P.C. Radlick, *Phytochemistry* 11, 1161 (1972).

Synthesis of 5'-deoxypyridoxal derivatives

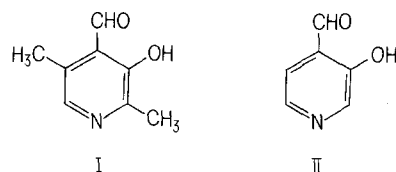
W. C. Cunningham and J. W. Thanassi^{1,2}

Department of Biochemistry, University of Vermont, College of Medicine, Burlington (Vermont 05405, USA), 27 September 1978

Summary. A convenient synthesis of 5'-deoxypyridoxal derivatives is described. The method involves catalytic hydrogenolysis of the corresponding 5'-phosphorylated derivatives; products are obtained in high yields.

5'-Deoxypyridoxal (**I**) and 3-hydroxypyridine-4-carboxaldehyde (**II**) are the best compounds available for the study of the mechanism of vitamin B_6 catalysis in model systems and have been used extensively for that purpose³⁻¹¹. These vitamin analogs have the functional groups essential for pyridoxal phosphate-like catalysis and at the same time avoid the complications associated with pyridoxal phosphate and pyridoxal. In the case of pyridoxal phosphate, kinetic and equilibrium studies as a function of pH become more difficult owing to ionizations of the 5'-phosphate group. In the case of pyridoxal, internal hemiacetal formation between the 5-hydroxymethyl group and the 4-carboxyaldehyde group creates a condition that does not exist in the enzymically active, phosphorylated form of the vitamin¹². Consequently, the vitamin analogs **I** and **II** are better in vitro model compounds than the naturally occurring forms of vitamin B_6 . In addition to model system studies, compounds of this type have been

used in the study of structure-activity relationships in vitamin B_6 -dependent enzymes¹³⁻¹⁶.



Several schemes for the synthesis of 5'-deoxypyridoxal, **I**¹⁷⁻¹⁹, and 3-hydroxypyridine-4-carboxaldehyde, **II**^{4, 20-22}, have been reported. However, even the best synthetic approach for the preparation of 5'-deoxypyridoxal requires 5 steps²³, starting from commercially available pyridoxine. We report in this communication a general synthetic method that conveniently leads, in high yields, to the